

Use of Saliva for Diagnosis of Diseases

J. Taaheri ¹, M. Bakhshi ², A. Aryankia ³, R. Noormohammadi ⁴✉

¹Professor, Department of Oral Medicine, School of Dentistry, Shahid Beheshti University of Medical Sciences, Tehran, Iran

²Assistant Professor, Department of Oral Medicine Department, School of Dentistry, Shahid Beheshti University of Medical Sciences, Tehran, Iran

³Postgraduate Student, Department of Periodontics, School of Dentistry, Shahid Beheshti University of Medical Sciences, Tehran, Iran

⁴Postgraduate Student, Department of Oral Medicine, School of Dentistry, Shahid Beheshti University of Medical Sciences, Tehran, Iran

Abstract

Background and Aim: Timely diagnosis is an important factor in successful treatment of diseases. In this regard, non-invasive sampling methods such as salivary analysis may be of great help in assessment of diseases and treatment outcomes.

Currently, introduction of new techniques for salivary analysis has increased its potential as a new diagnosing tool.

Materials and Methods: The National Institute of Dental and Craniofacial Research (NIDCR) was the first to propose the role of saliva in diagnosis and assessment of diseases. Since then, several studies have been conducted in this respect. This review article aims to evaluate the results of these studies to provide a brief insight on the correlation of saliva with different oral and systemic diseases. Key words namely saliva, biomarkers, oral diseases and systemic diseases were searched in PubMed, Google Scholar, EBSCO, Scopus and Medline databases and articles relevant to the use of saliva for evaluation and detection of various diseases published during 1950-2011 were collected.

Results: Salivary analysis increases the collaboration between dentists and physicians and can be used for detection of some systemic conditions.

Conclusion: This review study discusses the role of saliva in detection of different oral and systemic conditions and emphasizes its importance.

Key Words: Saliva, Biomarkers, Oral diseases, Systemic diseases

✉ Corresponding author:
R. Noormohammadi,
Postgraduate Student, Department of Oral Medicine, School of Dentistry, Shahid Beheshti University of Medical Sciences, Tehran, Iran
robab.noormohammadi@yahoo.com

Received: 8 Jan 2013
Accepted: 19 Dec 2013

Journal of Islamic Dental Association of IRAN (JIDAI) Spring 2014 ;26, (2)

Introduction

The main salivary glands are in the form of pairs and include the parotid, the submandibular and the sublingual glands. These glands secrete saliva into the oral cavity. The secreted saliva is composed of a combination of serous and mucosal components. A total amount of 1-1.5L of non-stimulated saliva is produced daily by these glands. The share of each gland in secretion is different, with the submandibular glands having 65%, parotid 23% and sublingual glands having 4% of the share; the minor salivary glands secrete only a small amount

(8%) of saliva [1]. In a more general sense, saliva can be divided into:

a) gland-specific saliva and

b) whole saliva [2]. Several factors affect the flow of secretion and the physiological characteristics of saliva. These factors include the circadian rhythm and physical activity [3]. Saliva is a clear, relatively acidic liquid (pH=6.0-7.0) containing electrolytes, immunoglobulins, proteins and different enzymes that play important roles in oral health. The primary roles of saliva include protecting the integrity of mucosal membrane, lubrication,

buffering and cleaning the oral cavity, antimicrobial effects, gustatory sense and primary digestion of food [1]. In the recent years, saliva has been extensively studied and the review of literatures shows that saliva can be used as a biological sample for diagnosis of diseases. The most important advantage of saliva analysis is its ease of use, accessibility and non-invasiveness. Additionally, it is possible to look for hormonal, microbiological, immunological, pharmacological and oncological markers in the saliva and compare their salivary concentration with their plasma concentration [4]. Many blood markers enter the saliva through the intercellular space and trans-cellular pathways (passive transmission and active transmission) or through paracellular pathways (extracellular ultrafiltration). Thus, the saliva reflects the physiological status of the body as efficiently as the plasma. Since the salivary concentration of these markers is less than their blood concentration, use of saliva for clinical diagnoses may not be sufficiently accurate [1]. Of course, the advent of new sensitive technologies such as molecular diagnostic techniques and nanotechnology has resolved this problem and proteins and salivary peptides are measured using different biochemical techniques like liquid chromatography, gel electrophoresis, capillary electrophoresis, nuclear magnetic resonance, immunoassay and Lectin probe analysis and they are used as diagnostic biomarkers [5].

Materials and Methods

The National Institute of Dental and craniofacial Research (NIDCR) was the first to introduce the role of saliva for diagnosis of diseases. Many previous studies have been conducted in this respect and the aim of this review article is to collect, classify and present a summary of the results of previous studies in such way that the reader can access brief and precise information on the correlation of different oral and systematic diseases with the composition of saliva. For this reason, key words described below were searched in Google, EBSCO, Scopus, Medline and PubMed databases and papers regarding the use of saliva for diagnosis of diseases published between 1950 and 2011 were collected. The searched key words included "saliva", "oral diseases" (dental caries, periodontal disease, lichen

planus, etc.) and "systemic diseases" (inherited, autoimmune, malignancies, etc.).

Dental Caries

Different researchers have tried to find a correlation between the composition of saliva and prevalence of dental caries. Recently, the salivary tests are commonly used for measuring certain oligosaccharides associated with dental caries [5]. Certain proteome changes in the saliva indicate dental caries. For example, proteins with high amounts of proline (PRP1 and PRP3), histatin 1 and statin decrease in high-caries patients while the level of these proteins is high in caries-free individuals. Histatin S and statin are useful predictors for caries [5,6,7]. Some studies have shown an association between the increased number of microorganisms like *Streptococcus mutans* and *Lactobacilli* in the saliva and dental caries [7]. The modified Saliva Check SM and Saliva Check sIgA are used for rapid and precise diagnosis of dental caries in the elderly and have been proven to be significantly useful [8]. In another study, physiochemical features like the flow of secretion, pH, the buffering capacity, calcium content, and protein and antioxidant level were compared in the saliva of children with and without dental caries. The results showed that the secretion of saliva, pH and the buffering capacity of saliva in children with dental caries were slightly lower; while the whole protein and anti-oxidant level were significantly higher than those in children without caries. The total calcium content of the saliva of children with caries was significantly lower than that in caries-free children [9].

Periodontal Disease

Existence of certain periopathogens in the saliva indicates that saliva is an appropriate means for the analysis of bacterial risk factors using DNA-based enzymes. This method is very useful to assess the course of periodontal disease [6]. Many studies have been performed using this method and different kinds of salivary proteins have been detected in periodontal disease [5]. Comparison of the salivary proteome of patients with invasive periodontitis and healthy subjects revealed that eleven salivary proteins such as α amylase, albumin and carbonic anhydrase 6 were different [5]. Previous studies have measured markers like matrix metalloproteinases (MMP), interleukins, hepatocellu-

lar growth factor (HGF), TNF- α and biomarkers related to bone turnover like the Macrophage Inflammatory Protein-1 α (MIP-1 α) and osteoprotegerin using different methods and reported an increase in the amount of these biomarkers in periodontal disease [10,11,12,13,14,15]. In another study, salivary calcitonin in subjects with type II diabetes reflected the severity of periodontitis and high blood sugar [16]. A significant increase in salivary level of TLR2 and 4 (toll like receptor) was found in patients affected with periodontitis. These factors enter the blood circulation from the gastrointestinal tract and increase the risk of atherosclerosis and insulin resistance [17]. Other studies have shown that high salivary concentration of PGE2 and TGF- β 2 in HIV⁺ patients is an important risk factor for progress of periodontitis [18, 19].

Oral Candidiasis

Saliva analysis is very efficient in diagnosis of oral mycosis. In oral candidiasis, fungi are colonized in the mucosa and make some changes in salivary proteomes. Proteins like calprotectin, histatin 5, mucin, peroxidase and high proline content proteins in the saliva have important diagnostic and prognostic roles in this regard [6]. In one study, a significant increase in salivary nitrates in these patients was observed [20]. Also, a reduction in antimicrobial proteins in the saliva causes an increase in candidiasis [21]. In another ex vivo study, the protective role of histatin 5 in the saliva in protection of oral mucosa was introduced for the first time. The results of previous studies have shown a significant decrease in level of histatin 5 in HIV⁺ positive patients with fungal colonization.

Lichen Planus

Salivary proteins are used as markers for detection of oral diseases. For example in patients with lichen planus, a decrease in salivary protein PLUNG (palate, lung and nasal epithelium carcinoma associated protein) was observed [5]. In one study, the salivary concentration of IL10, STNFR-2, INF- α and INF- δ in patients with erosive lichen planus was found to be high. The concentration of these markers decreased significantly after treatment with prednisone [24, 25]. In another study, level of IL-6, IL-1 α and TNF- α reduced significantly after treatment with dexamethasone and the results showed that the measurement of cytokines was useful in monitoring the response to treatment.

Burning Mouth Syndrome (BMS)

Saliva has also been used for evaluation of BMS. Although this syndrome is idiopathic, neuropathy has been suggested as a possible etiology. It means that there is an imbalance in substance P, Neurokinin A and calcitonin gene related peptide (CGRP) and the measurement of these neuropeptides in the saliva in comparison with serum showed that CGRP reduced in patients with BMS. However, these findings were in contrast to the results of Vanja et al, in 2010 [26, 27]. In another study, the amount of Nerve Growth Factor (NGF), substance P and products of degranulation of mast cells and neutrophils in the saliva was measured in the control and BMS patients. The results showed a significant increase in NGF and mast cell products, a significant decrease in SP and no change in neutrophil markers. Thus, these factors can be used for diagnosis and monitoring of BMS [29]. Also, in another study, a significant increase in salivary magnesium and no change in cytokines were noted after measuring and comparing the concentration of cytokines and markers in the saliva of healthy controls and BMS patients [30].

Recurrent Aphthous Stomatitis and Recurrent Aphthous Ulcer

Aphthous is one of the most common oral mucosal disorders with an unknown pathophysiology. Only a few factors are known to be responsible for its occurrence [31]. Researchers have measured the amounts of vitamins A, C and E and anti-oxidant MDA-5 (Malondialdehyde) in serum and saliva of patients with RAU in comparison with healthy controls and concluded that the amount of these vitamins was significantly lower in patients compared with healthy controls while the amount of MDA-5 was significantly higher in patients. Thus, there was a significant correlation between the saliva and serum of these patients [32]. Another study showed a reduction in salivary EGF (epithelial growth factor) in patients with high frequency of RAU [33]. This marker has also been measured in patients with RAS and Behcet's disease and the results, similar to the above-mentioned study, showed a decrease in the level of this factor. These findings indicate the role of EGF in pathogenesis of BD, RAS, RAU and mucosal disruption [34]. The results of another study showed a decrease in phagocytic function of salivary neutrophils similar to

blood neutrophils compared with healthy controls [35]. A similar study in this regard demonstrated that in the active phase of RAS, the salivary concentration of TNF- α is 2.5 times higher in patients than in healthy controls. Therefore, changes in TNF- α level have been suggested as the etiology of RAS [36]. Salivary and serum antioxidants have also been studied in patients with recurrent aphthous and the results have shown that changes in superoxide dismutase in red blood cells compared with other defensive substances like catalase and whole antioxidant play an important role in the inflammatory process seen in RAS [37]. The results of another study indicated that the concentration of salivary IgA in the active phases of RAS was significantly higher compared with the latent phases [38].

Systematic Diseases

Some systematic diseases affect the salivary glands either directly or indirectly and thus, change the components of the secreted saliva. Therefore, it is possible to use such changes for early diagnosis, determination of severity and even prognosis of diseases.

Hereditary Diseases Cystic Fibrosis

Cystic fibrosis is a genetic disorder that results from a mutation in the CFTR gene that is located on the long arm of chromosome 7 and mostly affects children and young adults. These people have general exocrinopathy and its recessive form is fatal. The abnormal secretion of exocrine glands in CF has led the clinicians to think about the efficacy of using saliva in diagnosis of this disease [39]. Increase in calcium, phosphorus, sodium, chloride, urea and uric acid in the saliva of these patients has been reported. The abnormal forms of Epithelial Growth Factor (EGF) and PGE-2 are also elevated in the saliva of these people. In addition, the activity of cathepsin D in these patients is significantly higher than in healthy controls. Salivary thiocyanate in these patients is also higher than in normal controls; although more studies are needed in this regard [6, 39, 40, 41, 42].

Celiac Disease

Celiac disease is a gastrointestinal disorder related to permanent gluten intolerance that causes the destruction of intestinal mucosa through immunologic mechanisms in people who have genetic predisposition. Atrophy of intestinal villi causes malab-

sorption of different types of nutrients like calcium, iron and folate. The results of several studies indicate that the amount of IgA Antigliadin Antibody (IgA-AGA) is elevated in the serum of these patients and this increase is in perfect alignment with salivary IgA-AGA. Therefore, it is possible to use this factor with a good sensitivity and specificity for screening of these patients. Some studies have even suggested the use of this test before performing intestinal villi biopsy [6, 39, 43, 44]. Other researchers have measured the anti-tissue transglutaminase immunoglobulin A (tTG-Abs) in the serum and saliva of patients and have concluded that screening and diagnosis of this disease using this factor in the saliva has adequate sensitivity and is simple. Moreover, the salivary tTG-Abs is detectable in these patients even when on a non-gluten diet [45, 46]. Another study on this topic has reported an increase in peroxidase, myeloperoxidase, albumin, total protein, IgA and IgG and a decrease in IgM and amylase in the saliva of these patients [47].

Deficiency or Lack of 21-Hydroxylase

Deficiency or lack of 21-hydroxylase is a hereditary steroidogenesis disorder that causes hyperplasia of the adrenal glands [48]. In the non-classic 21 hydroxylase disorder, there is partial deficiency of this enzyme. Primary monitoring of 17-dehydroxy progesterone of the saliva (17-DHP) is a perfect screening test for this disease because the salivary 17DHP reflects its concentration in serum [49, 50].

Sjögern's Syndrome (an autoimmune disorder)

Sjögern's syndrome (SS) is an autoimmune disorder that mainly affects functions of lacrimation and salivary glands and causes serum abnormalities. It can be detected by expensive and invasive tests. Recently, the use of saliva has been discussed in many studies for the diagnosis of this disease; which is quite valuable considering its non-invasiveness and affordable cost. Sialochemistry has been useful in diagnosis of Sjögern's syndrome. The results of saliva analysis in people with this disorder have shown an increase in inflammatory proteins, β -microglobulin, lactoferrin, lysozyme C, sodium, chloride, IgA, IgG, albumin and cystatins and decrease in high proline containing proteins and amylase. Also, there is a reduction in the saliva flow in SS; which is not a pathognomonic feature of this syndrome but will cause oral symptoms like caries,

fungal infections, dysphagia and oral pains [39, 51, 52]. Genomic biomarkers are also present in the saliva of these patients. In one study, 25 sequences of amino acids were identified to be up-regulated and 16 sequences were found to be down-regulated. Of these sequences, 10 up-regulated regions and 6 down-regulated regions were significantly useful for diagnosis of SS. Also, 162 types of mRNA were identified in the saliva that lead to interferon production, lymphocyte sequestration and antibody production involved in pathogenesis of SS [6]. In another study, the level of IL-7 in the saliva of minor glands in patients with SS was elevated. This increase can cause absorption and differentiation of T cells and lead to secretion of different cytokines like monokines, IL-4, IL-17, TNF- α and IL-1 α in the saliva of patients compared with healthy controls. Also, an increase in level of IL-18 (which is a pro-inflammatory and immunoregulatory cytokine and has a pathogenic role in autoimmune disease with Th1 involvement) was observed in serum and saliva of these patients that can indicate the involvement of this cytokine in the immunologic and inflammatory course of this disease. Proteomic analysis has also been used for distinction of primary and secondary Sjögren's syndrome [53, 54, 55, 56].

Malignancies

Early detection and treatment of cancers is important in increasing life expectancy. Certain markers in the serum, saliva and other body fluids can be useful in detection of cancers. Thus, some studies have used proteomic and genomic analyses for diagnosis and prognosis of cancers. Evidence shows that mutation of P53 gene is detectable in the salivary DNA of 62.5% of patients with different oral cancers. Also, HPV virus DNA increases in head and neck cancers. Studies related to salivary mRNA profiling show 4 major biomarkers in the saliva of patients with oral cancer [IL-1 β , IL-8, ornithine decarboxylase antizyme-1 (OAZ1) and spermidine N1 acetyltransferase (SAT)]. Salivary proteome has also shown an increase in defensin-1 and cancerous antigens CA 15-3, CA-125 and P53 in different cancers [6, 39, 57, 59, 58]. Increased levels of IL-6 and FGF- β have been found in the saliva and serum of oral SCC patients. Such increase probably indicates that the origin of these factors is the carcinomatous cells. Another study

measured the amount of IL-1, IL-6, IL-8 and TNF- α in patients with oral SCC as well as patients with pre-cancerous disease in comparison with healthy controls. The results showed different levels of these cytokines in OSCC and OPML; therefore, these pro-angiogenic and pro-inflammatory cytokines can be used for diagnostic and prognostic purposes [60, 61, 62]. In another study the amount of salivary endothelin-1 in oral SCC and leukoplakia was studied and the results indicated that the amount of this factor was elevated in OSCC; but it was not a good marker for evaluation of the degree of malignancy [64]. Other studies detected a significant increase in different markers like IL-6 and IL-8 and also salivary glutathione of patients with oropharyngeal SCC compared with healthy controls [65, 66]. In one study, a significant increase in the concentration of different MMP like MMP-7, MMP-9 and MMP-26 was detected in patients with SCC of the lower lip and tongue. Thus, MMP can be used as a diagnostic biomarker in this cancer [67]. In another study, the amount of magnesium, calcium, copper, chloride, potassium, sodium, total protein and amylase was measured and compared between OSCC patients and healthy controls. The results showed higher levels of salivary sodium and chloride and lower level of salivary total protein in these patients compared with controls [68]. Moreover, a similar study examined changes in 8 salivary biomarkers in patients with cancer and reported an increase in 5 markers (MMP, Ki 67, cyclin D1, lactate dehydrogenase and carbonyl) and a decrease in 3 markers (Maspin, phosphorylated-src and 8-oxoguanine-DNA). Since these markers have high sensitivity and specificity, they can be very useful in diagnosis of cancers in other parts of the body. For example, prostate specific antigen (PSA) in the saliva that is in complete correlation with blood PSA is useful for diagnosis of prostate adenocarcinoma [70]. Additionally, transcriptomic biomarkers in the saliva with high specificity and sensitivity have a good application in diagnosis of pancreatic cancer [71]. On the same topic, a study examined the transcriptomic and proteomic biomarkers in patients with breast cancer and healthy controls not influenced with confounding factors. The results revealed that these markers had high sensitivity and specificity for diagnosis of breast cancer [72]. In other studies, there was a significant increase in

factors like EGF, C-erb β 2, CA-125, glycoprotein and CA 15-3 in the saliva of patients with breast and ovarian cancers [58, 59, 73, 74, 75, 76, 77].

Endocrinology

Collection of saliva facilitates determination of hormonal level and diurnal variation [78]. Studies have shown that salivary cortisol is a valuable indicator of serum cortisol which is not dependent on salivary flow. Indeed, clinical assessment of most endocrine glands requires monitoring of plasma steroids. It has been found that while the amount of steroid hormones in the saliva reflects the free and active amounts of these hormones, the blood shows its total amount (free and bound) [76]. Different studies have evaluated salivary levels of cortisol, estradiol, progesterone and testosterone. Since these hormones play an important role in mental health of patients, they can also be used in the field of psychology. It has been shown that testosterone level in women with breast cancer is significantly low [79]. In another study, the salivary levels of progesterone and estradiol were assessed in pregnant women and the increase of estradiol was found to increase the risk of premature labor [80, 81]. The amount of salivary cortisol in other studies indicated the level of serum cortisol. Based on the literature, the morning concentration of salivary cortisol can be used for the diagnosis of Addison's disease and its nocturnal concentration can be used for diagnosis of Cushing's disease [82, 83, 84, 85]. Another study assessing the seasonal changes of salivary testosterone concentration concluded that the concentration of this hormone increased in autumn and decreased in summer. These results can now be used for research in the field of psychology and endocrinology [86]. Exercise also changes the salivary concentration of steroid hormones, immunoglobulins, antimicrobial proteins and enzymes. The level of these hormones increases in irregular and unprofessional activities and decreases in professional sportsmen [87]. Likewise, it is possible to use cortisol concentration in non-stimulated saliva early in the morning for screening of patients predisposed to secondary adrenal insufficiency [88]. Beside these applications, the saliva can also be used for measurement of aldosterone [89, 90].

Diabetes

Different studies have shown that treatment-resistant periodontitis can be an indicator of undiag-

nosed or uncontrolled type 2 diabetes. Thus, resistant periodontal disease is a risk factor for cardiovascular diseases like diabetes [78]. On the other hand, certain salivary biomarkers are helpful in diagnosis of type 2 diabetes [91]. One study showed a significant increase in the activity of proteases like cathepsin D and MMP-9 in healthy controls and people with diabetes in such way that collagen fragments can be used for monitoring of pathologies related to diabetes [92]. The results of another study showed a significant increase in salivary IL-6 in diabetic patients who also had lichen planus compared with other groups (healthy controls, diabetics and people with lichen planus) [93]. In another study, use of saliva containing immunologic factors in people with diabetes led to a reduction in plaque, gingivitis and fungal diseases [94]. The analysis of saliva in another study showed an increase in salivary glucose and a decrease in salivary amylase of diabetic patients (controlled and uncontrolled) [95].

Renal and Cardiovascular Diseases

Saliva has also been used for detection of renal diseases. Some studies measured the salivary creatinine level and predicted the renal disease with high sensitivity and specificity [75].

B2-microglobulin is a major component in dialysis-related amyloidosis. In one study the amount of this marker was evaluated in the saliva and serum of healthy subjects, diabetics, patients with chronic renal disease and subjects under hemodialysis. The results showed that analysis of salivary β 2-microglobulin was a reliable method for assessing its serum concentration and can be used to predict the risk of amyloidosis, secondary to chronic renal disease [96]. Hyperphosphatemia is an important factor in vascular calcification in people with chronic renal failure. This calcification is the main cause of mortality in patients under hemodialysis and needs to be treated. The salivary phosphate after being swallowed also contributes to this problem. Since salivary phosphate is in perfect accordance with serum phosphate, it can be a good marker for initiation of treatment in this process [97]. Another study used salivary urea and nitrogen which are in imperfect accordance with blood urea and nitrogen for detection of chronic renal failure [98]. The results of a similar study indicated that the salivary concentration of creatinine, urea, sodium,

potassium, chloride and α -amylase in patients with chronic renal failure was higher than in the control group while the calcium level was significantly lower. It should be noted that the components of saliva vary with the severity of disease [99]. Other studies have shown a significant decrease in salivary concentration of inflammatory cytokines in patients with chronic renal failure [100]. Moreover, saliva can be used for monitoring of cardiovascular diseases. Currently, cardiovascular diseases are a major cause of mortality. Use of salivary markers for monitoring of these patients has been confirmed especially after a cardiac surgery. An example of these markers is the salivary α -amylase; which is elevated in stressful situations [6, 76].

Also, periodontal disease by increasing IL-6, TNF- α and C-reactive protein (CRP) plays an important role in risk determination of cardio-metabolic diseases. Biomarkers like myoglobin, myeloperoxidase and CRP have a significant diagnostic ability in myocardial infarction [78] and the amount of collagenases like MMP increases in cardiovascular diseases [13].

Other applications: Many studies have been performed on the efficacy of saliva in diagnosis of different viral and bacterial diseases and monitoring of different medications; which will not be discussed here due to the limitations of the current study.

Currently, there are many studies that have assessed different biomarkers (genomic, transcriptomic and proteomic) and found their correlation with some diseases. Specific kits are being designed for measuring salivary markers in different diseases. By doing so, a fast, cost effective and easy method will replace the existing ones. Using this test we can measure diagnostic markers; which will help in early detection of different diseases.

Researchers have recently introduced a kit for measuring CRP; which is a general indicator of inflammatory diseases [101].

It appears that with recent advancements in technology, saliva analysis will soon be a valuable method for diagnosis.

References

1. Lee YH, Wong DT. Saliva: An emerging biofluid for early detection of diseases. *Am J Dent*. 2009 Aug; 22(4):241-8.
2. Kaufman E, Lamster IB. The diagnostic applications of saliva--a review. *Crit Rev Oral Biol Med*. 2002 March; 13(2):197-212.
3. Dawes C. Considerations in the development of diagnostic tests on saliva. *Ann N Y Acad Sci*. 1993 Sept 20; 694:265-9.
4. Lee JM, Garon E, Wong DT. Salivary diagnostics. *Orthod Craniofac Res*. 2009 Aug; 12(3):206-11.
5. Kawas SA, Rahim ZH, Ferguson DB. Potential uses of human salivary protein and peptide analysis in the diagnosis of disease. *Arch Oral Biol*. 2012 Jul; 57(1):1-9.
6. Fábán TK, Fejérdy P, Csermely P. Salivary Genomics, transcriptomics and proteomics: The Emerging Concept of the Oral Ecosystem and their use in the early diagnosis of cancer and other diseases. *Curr Genomics*. 2008 Mar; 9(1):11-21.
7. Rudney JD, Staikov RK, Johnson JD. Potential biomarkers of human salivary function: a modified proteomic approach. *Arch Oral Biol*. 2009 Jan; 54(1):91-100.
8. Senpuku H, Miyazaki H, Yoneda S, Yoshihara A, Tada A. A quick statistically accurate diagnosis for caries risk in the elderly. *Clin Lab*. 2010; 56(11-12):505-12.
9. Preethi BP, Reshma D, Anand P. Evaluation of Flow Rate, pH, buffering capacity, calcium, total proteins and total antioxidant capacity levels of saliva in caries free and caries active children: An In Vivo Study. *Indian J Clin Biochem*. 2010 Oct; 25(4):425-8.
10. Miller CS, King CP Jr, Langub MC, Kryscio RJ. Salivary biomarkers of existing periodontal disease: A cross-sectional study. *J Am Dent Assoc*. 2006 Mar; 137(3):322-9.
11. Rai B, Kharb S, Jain R, Anand SC. Biomarkers of periodontitis in oral fluids. *J Oral Sci*. 2008 Mar; 50(1):53-612.
12. Al-Sabbagh M, Alladah A, Lin Y, Kryscio RJ. Bone remodeling-associated salivary biomarker MIP-1 α distinguishes periodontal disease from health. *J Periodontal Res*. 2012 June; 47(3):389-395.
13. Collagenase-2 (MMP-8) as a point-of-care biomarker in periodontitis and cardiovascular diseases. Therapeutic response to non-antimicrobial properties of tetracyclines. *Pharmacol Res*. 2011 Feb; 63(2):108-13.

14. Sorsa T, Tervahartiala T, Leppilähti J, Hernandez M. Significance of elevated gingival crevicular fluid tumor necrosis factor- α and interleukin-8 levels in chronic hemodialysis patients with periodontal disease. *J Periodontol Res.* 2010 Aug; 45(4):445-50.
15. Rudrakshi C, Srinivas N, Mehta DS. A comparative evaluation of hepatocyte growth factor levels in gingival crevicular fluid and saliva and its correlation with clinical parameters in patients with and without chronic periodontitis: A clinico-biochemical study. *J Indian Soc Periodontol.* 2011 Apr; 15(2):147-51.
16. Bassim CW, Redman RS, DeNucci DJ, Becker KL. Salivary procalcitonin and periodontitis in diabetes. *J Dent Res.* 2008 Jul; 87(7):630-4.
- Faculty of dentistry Jundi Shapur University Of
17. Lappin DF, Sherrabeh S, Erridge C. Stimulants of Toll-like receptors 2 and 4 are elevated in saliva of periodontitis patients compared with healthy subjects. *J Clin Periodontol.* 2011 Apr; 38(4):318-25.
18. Alpagot T, Remien J, Bhattacharyya M, Konopka K. Longitudinal evaluation of prostaglandin E₂ (PGE₂) and periodontal status in HIV+ patients. *Arch Oral Biol.* 2007 Nov; 52(11):1102-8.
19. Alpagot T, Konopka K, Bhattacharyya M, Gebremedhin S. The association between gingival crevicular fluid TGF- β 1 levels and periodontal status in HIV-1(+) patients. *J Periodontol.* 2008 Jan; 79(1):123-30.
20. Shi RT, Qin LZ, Xia DS, Deng DJ, Fan ZP, Shan ZC, Xu YY, Wang SL. Increase of saliva nitrate and nitrite level in patients with oral candidiasis. *Zhonghua Yu Fang Yi Xue Za Zhi.* 2009 Jul; 43(7):607-10.
21. Toyohiro Tanida¹, Tetsuro Okamoto¹, Atsuko Okamoto¹, Haiyan Wang². Decreased excretion of antimicrobial proteins and peptides in saliva of patients with oral candidiasis. *J Oral Pathol&Med.* 2003 Nov; 32(10):586-594.
22. Brian M, Peters, Jingsong Zhu, Paul L. Protection of the oral mucosa by salivary histatin-5 against *Candida albicans* in an ex vivo murine model of oral infection. *FEMS Yeast Res.* 2010 Aug;10(5):597-604.
23. Sandra R, Torres, Alfredo Garzino-Demo, Timothy F. Salivary histatin-5 and oral fungal colonisation in HIV+ individuals. *Mycoses.* 2009 Jan; 52(1):11-15.
24. Ghallab NA, el-Wakeel N, Shaker OG. Levels of salivary IFN- γ , TNF- α , and TNF receptor-2 as prognostic markers in (erosive) oral lichen planus. *Mediators Inflamm.* 2010 Feb;847632.
25. Dan H, Liu W, Wang J, Wang Z. Elevated IL-10 concentrations in serum and saliva from patients with oral lichen planus. *Quintessence Int.* 2011 Feb; 42(2):157-63.
26. Rhodus NL, Cheng B, Bowles W, Myers S. Proinflammatory cytokine levels in saliva before and after treatment of (erosive) oral lichen planus with dexamethasone. *Oral Dis.* 2006 Mar; 12(2):112-6.
27. Zidverc-Trajkovic J, Stanimirovic D, Obrenovic R, Tajti J. Calcitonin gene-related peptide levels in saliva of patients with burning mouth syndrome. *J Oral Pathol Med.* 2009 Jan; 38(1):29-33.
28. Boras VV, Savage NW, Brailo V, Lukac J, Lukac M, Alajbeg IZ. Salivary and serum levels of substance P, neurokinin A and calcitonin gene related peptide in burning mouth syndrome. *Med Oral Patol Oral Cir Bucal.* 2010 May; 15(3):e427-31.
29. Borelli V, Marchioli A, Di Taranto R, Romano M, Chiandussi S, Di Lenarda R, Biasotto M, Zambucchi G. Neuropeptides in saliva of subjects with burning mouth syndrome: A pilot study. *Oral Dis.* 2010 May; 16(4):365-74.
30. Pekiner FN, Gümrü B, Demirel GY, Ozbayrak S. Burning mouth syndrome and saliva: Detection of salivary trace elements and cytokines. *J Oral Pathol Med.* 2009 Mar; 38(3):269-75.
31. Zunt SL. Recurrent aphthous stomatitis. *Dermatol Clin.* 2003 Jan;21(1):33-9.
32. Saral Y, Coskun BK, Ozturk P, Karatas F, Ayar A. Assessment of salivary and serum antioxidant vitamins and lipid peroxidation in patients with recurrent aphthous ulceration. *Tohoku J Exp Med.* 2005 Aug;206(4):305-12.
33. Gu Y, Zhang G, Lin M. Quantity research on epidermal growth factor in saliva and epidermal growth factor receptor in biopsy samples of recurrent aphthous ulcer patients. *Hua Xi Kou Qiang Yi Xue Za Zhi.* 2008 Feb; 26(1):36-9.
34. Adışen E, Aral A, Aybay C, Gürer MA. Salivary epidermal growth factor levels in Behçet's

disease and recurrent aphthous stomatitis. *Dermatol.* 2008Jul; 217(3):235-40.

35. Kumar BP, Keluskar V, Bagewadi AS, Shetti A. Evaluating and comparing phagocytic functions of salivary and blood neutrophils in patients with recurrent aphthous ulcers and controls. *Quintessence Int.* 2010 May;41(5):411-6.

36. Eguia-del Valle A, Martinez-Conde-Llamas R, López-Vicente J. Salivary levels of tumour necrosis factor-alpha in patients with recurrent aphthous stomatitis. *Med Oral Patol Oral Cir Bucal.* 2011 Jan; 16(1):e33-6.

37. Momen-Beitollahi J, Mansourian A, Momen-Heravi F, Amanlou M, et al. Assessment of salivary and serum antioxidant status in patients with recurrent aphthous stomatitis. *Med Oral Patol Oral Cir Bucal.* 2010 Jul; 15(4):e557-61.

38. Martinez Kde O, Mendes LL, Alves JB. Secretory A immunoglobulin, total proteins and salivary flow in Recurrent Aphthous Ulceration. *39 Braz J Otorhinolaryngol.* 2007 May-Jun; 73(3):323-8.

39. Kaufman E, Lamster IB. The diagnostic applications of saliva-a review. *Crit Rev Oral Biol Med.* 2002 Jan; 13(2):197-212.

40. Minarowska A, Minarowski L, Karwowska A, Sands D, Dabrowska E. The activity of cathepsin D in saliva of cystic fibrosis patients. *Folia Histochem Cytobiol.* 2007 July;45(3):165-8.

41. Minarowski L, Sands D, Minarowska A, Karwowska A, Sulewska A, Gacko M, Chyczewska E. Thiocyanate concentration in saliva of cystic fibrosis patients. *Folia Histochem Cytobiol.* 2008April; 46(2):245-6.

42. Minarowski L, Minarowska L, Karwowska A. Activity of lysosomal carboxypeptidase A in mixed saliva of cystic fibrosis patients. *J Cystic Fibrosis.* 2008Jun;7(2):1-34.

43. Rashid M, Zarkadas M, Anca A, Limeback H. Oral manifestations of celiac disease: A clinical guide for dentists. *J Can Dent Assoc.* 2011Oct; 77: b25-39.

44. Hakeem V, Fifield R, al-Bayaty HF, Aldred MJ. Salivary IgA antigliadin antibody as a marker for coeliac disease. *Arch Dis Child.* 1992 Jun; 67(6):724-7.

45. Bonamico M, Nenna R, Montuori M, Luparia RP. First salivary screening of celiac disease by detection of anti-transglutaminase autoantibody radioimmunoassay in 5000 Italian primary school-

children. *J Pediatr Gastroenterol Nutr.* 2011 Jan; 52(1):17-20.

46. Bonamico M, Nenna R, Luparia RP, Perricone C. Radioimmunological detection of anti-transglutaminase autoantibodies in human saliva: A useful test to monitor coeliac disease follow-up. *Aliment Pharmacol Ther.* 2008 Aug1;28(3):364-70.

47. Lenander-Lumikari M, Ihalin R, Lähteenoja H. Changes in whole saliva in patients with coeliac disease. *Arch Oral Biol.* 2000 May; 45(5):347-54.

48. Concolino P, Mello E, Zuppi C, Capoluongo E. Molecular diagnosis of congenital adrenal hyperplasia due to 21-hydroxylase deficiency: an update of new CYP21A2 mutations. *Clin Chem Lab Med.* 2010 Aug; 48(8):1057-62.

49. Zerah M, Ueshiba H, Wod E, Speiser PW. Prevalence of nonclassical steroid 21-hydroxylase deficiency based on a morning salivary 17-hydroxyprogesterone screening test: A small sample study. *J Clin Endocrinol Metab.* 1990 Jun; 70(6):1662-7.

50. Zerah M, Pang SY, New MI. Morning salivary 17-hydroxyprogesterone is a useful screening test for nonclassical 21-hydroxylase deficiency. *J Clin Endocrinol Metab.* 1987 Aug; 65(2):227-32.

51. Hu S, Wang J, Meijer J, Jeong S, Xie Y, Yu T, Zhou H, Henry S, Vissink A, Pijpe J, Kallenberg C, Elashoff D, Loo JA, Wong DT. Salivary proteomic and genomic biomarkers for primary Sjögren's syndrome. *Arthritis Rheum.* 2007 Nov; 56(11):3588-600.

52. Ryu OH, Atkinson JC, Hoehn GT, Illei GG. Identification of parotid salivary biomarkers in Sjogren's syndrome by surface-enhanced laser desorption/ionization time-of-flight mass spectrometry and two-dimensional difference gel electrophoresis. *Rheumatology (Oxford).* 2006 Sept;45(9): 1077-86.

53. Bikker A, van Woerkom JM, Kruize AA, Wenting-van Wijk M. Increased expression of interleukin-7 in labial salivary glands of patients with primary Sjögren's syndrome correlates with increased inflammation. *Arthritis Rheum.* 2010 Apr; 62(4):969-77.

54. Bombardieri M, Barone F, Pittoni V, Alessandri C. Increased circulating levels and salivary gland expression of interleukin-18 in patients with Sjögren's syndrome: Relationship with autoantibody production and lymphoid organization of the

- periductal inflammatory infiltrate. *Arthritis Res Ther*. 2004 Aug; 6(5):R447-56.
55. Baldini C, Giusti L, Bazzichi L, Lucacchini A. Proteomic analysis of the saliva: A clue for understanding primary from secondary Sjögren's syndrome? *Autoimmun Rev*. 2008 Jan; 7(3):185-91.
 56. Ferraccioli G, De Santis M, Peluso G, Inzitari R. Proteomic approaches to Sjögren's syndrome: A clue to interpret the pathophysiology and organ involvement of the disease. *Autoimmun Rev*. 2010 Jul; 9(9):622-6.
 57. Nagler RM. Saliva as a tool for oral cancer diagnosis and prognosis. *Oral Oncol*. 2009 Dec; 45(12):1006-10.
 58. Agha-Hosseini F, Mirzaei-Dizgah I, Rahimi A. Correlation of serum and salivary CA15-3 levels in patients with breast cancer. *Med Oral Patol Oral Cir Bucal*. 2009 Oct 1;14(10):e521-4.
 59. Agha-Hosseini F, Mirzaei-Dizgah I, Rahimi A, Seilanian-Toosi M. Correlation of serum and salivary CA125 levels in patients with breast cancer. *J Contemp Dent Pract*. 2009 Nov 1;10(6):E001-8.
 60. Rhodus NL, Ho V, Miller CS, Myers S, Ondrey F. NF-kappaB dependent cytokine levels in saliva of patients with oral preneoplastic lesions and oral squamous cell carcinoma. *Cancer Detect Prev*. 2005 Jan;29(1):42-5.
 61. Wu JY, Yi C, Chung HR, Wang DJ, Chang WC, Lee SY, Lin CT, Yang YC, Yang WC. Potential biomarkers in saliva for oral squamous cell carcinoma. *Oral Oncol*. 2010 Apr; 46(4):226-31.
 62. Vucicević Boras V, Cikes N, Lukać J, Virag M, Cekić-Arambasin A. Salivary and serum interleukin 6 and basic fibroblast growth factor levels in patients with oral squamous cell carcinoma. *Minerva Stomatol*. 2005 Oct; 54(10):569-73.
 63. Hoffmann RR, Yurgel LS, Campos MM. Evaluation of salivary endothelin-1 levels in oral squamous cell carcinoma and oral leukoplakia. *Regul Pept*. 2011 Jan 17;166(1-3):55-8.
 64. Pickering V, Jordan RC, Schmidt BL. Elevated salivary endothelin levels in oral cancer patients-a pilot study. *Oral Oncol*. 2007 Jan; 43(1):37-41.
 65. St John MA, Li Y, Zhou X, Denny P, Ho CM, Montemagno C, Shi W, Qi F, Wu B, Sinha U, Jordan R, Wolinsky L, Park NH, Liu H, Abemayor E, Wong DT. Interleukin 6 and interleukin 8 as potential biomarkers for oral cavity and oropharyngeal squamous cell carcinoma. *Arch Otolaryngol Head Neck Surg*. 2004 Aug; 130(8):929-35.
 66. Almadori G, Bussu F, Galli J, Limongelli A, Persichilli S, Zappacosta B, Minucci A, Paludetti G, Giardina B. Salivary glutathione and uric acid levels in patients with head and neck squamous cell carcinoma. *Head Neck*. 2007 Jul; 29(7):648-54.
 67. Barros SS, Henriques AC, Pereira KM, de Medeiros AM, Galvão HC, Freitas Rde A. Immunohistochemical expression of matrix metalloproteinases in squamous cell carcinoma of the tongue and lower lip. *Arch Oral Biol*. 2011 Aug; 56(8):752-60.
 68. Fuchs PN, Rogić D, Vidović-Juras D, Susić M, Milenović A, Brailo V, Boras VV. Salivary analytes in patients with oral squamous cell carcinoma. *Coll Antropol*. 2011 Jun;35(2):359-62.
 69. Shpitzer T, Hamzany Y, Bahar G, Feinmesser R, Savulescu D, Borovoi I, Gavish M, Nagler RM. Salivary analysis of oral cancer biomarkers. *Br J Cancer*. 2009 Oct 6; 101(7):1194-8.
 70. Shiiki N, Tokuyama S, Sato C, Kondo Y, Saruta J, Mori Y, Shiiki K, Miyoshi Y, Tsukinoki K. Association between saliva PSA and serum PSA in conditions with prostate adenocarcinoma. *Biomarkers*. 2011 Sept; 16(6):498-503.
 71. Zhang L, Farrell JJ, Zhou H, Elashoff D, Akin D, Park NH, Chia D, Wong DT. Salivary transcriptomic biomarkers for detection of resectable pancreatic cancer. *Gastroenterology*. 2010 Mar; 138(3):949-57.e1-7.
 72. Zhang L, Xiao H, Karlan S, Zhou H, Gross J, Elashoff D, Akin D, Yan X, Chia D, Karlan B, Wong DT. Discovery and preclinical validation of salivary transcriptomic and proteomic biomarkers for the non-invasive detection of breast cancer. *Plos One*. 2010 Dec; 31;5(12):e15573.
 73. The use of soluble, salivary c-erbB-2 for the detection and post-operative follow-up of breast cancer in women: the results of a five-year translational research study. *Adv Dent Res*. 2005 Jun; 18(1):17-24.
 74. Streckfus C, Bigler L. The use of soluble, salivary c-erbB-2 for the detection and post-operative follow-up of breast cancer in women:

the results of a five-year translational research study. *Adv Dent Res.* 2005 Jun;18(1):17-24.

75. Chen DX, Li FQ. Primary research on saliva and serum CA125 assays for detecting malignant ovarian tumors. *Zhonghua Fu Chan Ke Za Zhi.* 1990 Mar;25(2):84-5, 123-4.
76. Streckfus CF, Bigler LR. Saliva as a diagnostic fluid. *Oral Dis.* 2002 Mar;8(2):69-76.
77. Pink R, Simek J, Vondrakova J, Faber E, Michl P, Pazdera J, Indrak K. Saliva as a diagnostic medium. *Biomed Pap Med Fac Univ Palacky Olomouc Czech Rep.* 2009 Jun; 153(2): 103-10.
78. Tremblay M, Gaudet D, Brisson D. Metabolic syndrome and oral markers of cardiometabolic risk. *J Can Dent Assoc.* 2011 Sept; 77:b125.
79. Dimitrakakis C, Zava D, Marinopoulos S, Tsigginou A, Antsaklis A, Glaser R. Low salivary testosterone levels in patients with breast cancer. *BMC Cancer.* 2010 Oct; 11; 10: 547.
80. Klebanoff MA, Meis PJ, Dombrowski MP. Salivary progesterone and estriol among pregnant women treated with 17-alpha-hydroxyprogesterone caproate or placebo. *Am J Obstet Gynecol.* 2008 Nov; 199(5): 506.e1-7.
81. Heine RP, McGregor JA, Goodwin TM, Artal R, Hayash RH, Robertson PA, et al. Serial salivary estriol to detect an increased risk of pre-term birth. *Obstet Gynecol.* 2000 Oct; 96(4): 490-7.
82. Deutschbein T, Unger N, Hinrichs J, Walz MK, Mann K, Petersenn S. Late-night and low-dose dexamethasone-suppressed cortisol in saliva and serum for the diagnosis of cortisol-secreting adrenal adenomas. *Eur J Endocrinol.* 2009 Nov; 161(5):747-53.
83. Carroll T, Raff H, Findling JW. Late-night salivary cortisol measurement in the diagnosis of cushing's syndrome. *Nat Clin Pract Endocrinol Metab.* 2008 Jun;4(6):344-50.
84. Perogamvros I, Keevil BG, Ray DW, Trainer PJ. Salivary cortisone is a potential biomarker for serum free cortisol. *J Clin Endocrinol Metab.* 2010 Nov;95(11):4951-8.
85. Restituto P, Galofré JC, Gil MJ, Mugueta C, Santos S, Monreal JJ, Varo N. Advantage of salivary cortisol measurements in the diagnosis of glucocorticoid related disorders. *Clin Biochem.* 2008 Jun;41(9):688-92.
86. Stanton SJ, Mullette-Gillman OA, Huettel SA. Seasonal variation of salivary testosterone in men, normally cycling women, and women using hormonal contraceptives. *Physiol Behav.* 2011 Oct 24; 104(5):804-8.
87. Papacosta E, Nassis GP. Saliva as a tool for monitoring steroid, peptide and immune markers in sport and exercise science. *J Sci Med Sport.* 2011 Sept; 14(5):424-34.
88. Deutschbein T, Unger N, Mann K, Petersenn S. Diagnosis of secondary adrenal insufficiency: unstimulated early morning cortisol in saliva and serum in comparison with the insulin tolerance test. *Horm Metab Res.* 2009 Nov; 41(11):834-9.
89. Manolopoulou J, Gerum S, Mulatero P, Rossignol P, Plouin PF, Reincke M, Bidlingmaier M. Salivary aldosterone as a diagnostic aid in primary aldosteronism. *Horm Metab Res.* 2010 Jun; 42(6):400-5.
90. Manolopoulou J, Mulatero P, Maser-Gluth C, Rossignol P, Spyroglou A, Vakrilova Y, et al. Saliva as a medium for aldosterone measurement in repeated sampling studies. *Steroids.* 2009 Oct; 74(10-11):853-8.
91. Rao PV, Reddy AP, Lu X, Dasari S. Proteomic identification of salivary biomarkers of type-2 diabetes. *96 J Proteome Res.* 2009 Jan; 8(1): 239-45.
92. Caseiro A, Vitorino R, Barros AS, Ferreira R. Salivary peptidome in type 1 diabetes mellitus. *Biomed Chromatogr.* 2012 May;26(5):571-82.
93. Liu Y, Jin JQ, Yuan ZF, Liu XS. Levels of interleukin-6 and tumor necrosis factor- α in saliva of patients with type 2 diabetes mellitus and oral lichen planus. *Beijing Da Xue Xue Bao.* 2011 Aug 18; 43(4):596-9.
94. Montaldo L, Montaldo P, Papa A, Caramico N. Effects of saliva substitutes on oral status in patients with Type 2 diabetes. *Diabet Med.* 2010 Nov; 27(11):1280-3.
95. Panchbhavi AS, Degwekar SS, Bhowte RR. Estimation of salivary glucose, salivary amylase, salivary total protein and salivary flow rate in diabetics in India. *J Oral Sci.* 2010 Sept; 52(3): 359-68.
96. Michelis R, Sela S, Ben-Zvi I, Nagler RM. Salivary beta2-microglobulin analysis in chronic

kidney disease and hemodialyzed patients. *Blood Purif.* 2007 Jan; 25(5-6):505-9.

97. Savica V, Calò L, Santoro D, Monardo P, Granata A, Bellinghieri G. Salivary phosphate secretion in chronic kidney disease. *J Ren Nutr.* 2008 Jan; 18(1):87-90.

98. Raimann JG, Kirsits W, Gebetsroither E, Carter M, Callegari J, Rosales L, et al. Saliva urea dipstick test: application in chronic kidney disease. *Clin Nephrol.* 2011 Jul; 76(1):23-8.

99. Tomás I, Marinho JS, Limeres J, Santos MJ. Changes in salivary composition in patients with renal failure. *Arch Oral Biol.* 2008 Jun; 53(6): 528-32.

100. Thorman R, Lundahl J, Yucel-Lindberg T, Hylander B. Inflammatory cytokines in saliva: early signs of metabolic disorders in chronic kidney disease. A controlled cross-sectional study. *Oral Surg Oral Med Oral Pathol Oral Radiol Endod.* 2010 Nov; 110(5):597-604.

101. Farnaud SJ, Kostic O, Getting SJ, Renshaw D. Saliva: Physiology and diagnostic potential in health and disease. *Scientific World J.* 2010 Mar 16; 10:434-56.